# A Novel Strategy in HIV Eradication using CD8 CAR T cells

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# INTRODUCTION

Latent HIV reservoir are a major burden for HIV cure as they are not recognised and killed by CD8 T cells. Guiding CD8 T cells to target these cells can be achieved by engineering chimeric antigen receptor (CAR) CD8 T cells using HIV broadly neutralising antibodies (bNAbs).

Accessing infected tissue resident CD4 memory T cells  $(T_{RM})$  provides additional challenge in curing HIV. As most  $T_{RM}$  express CD69, incorporating CD69 ligand, Galectin-1 (Gal-1), into CD8 CAR T cells might help direct the CAR T cells into tissue.

Having a low-cost and high genetic capacity, *piggyBac* transposon system was used to deliver CAR genes via electroporation to generate CD8 CAR T cells.

# HYPOTHESIS

1) CD8 CAR T cells will have specific activation and cytotoxicity against *in vitro* HIV model

2) Targeting HIV antigens combined with CD69 will be a novel strategy to eradicate latently infected CD4 T cells within tissues

# AIMS

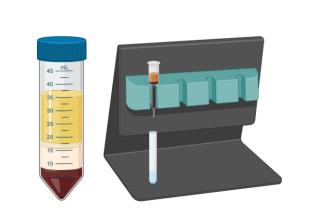
1) Generate and test single bNAb (PGT121, VRC07)-

based and Galectin-1 (Gal-1) CD8 CAR T cells

2) Generate and test Split Tandem CD8 CAR (TanCAR) T cells expressing Gal-1 and two bNAbs utilising AND-gated strategy

## METHOD

# Isolate CD8 T cells



Transfection of CD8 T cells

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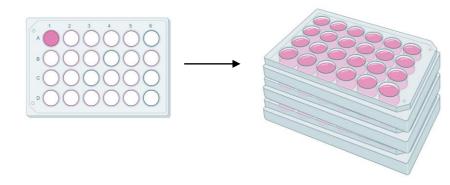
Day 1

piggyBac transposon delivered via electroporation:

5x10<sup>6</sup> cells

**Day 1-21** 

CD8 CAR T cells Stimulation and Expansion

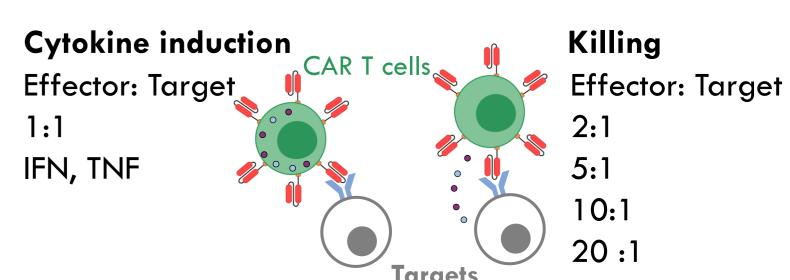


400 IU IL15 (every 2-3 days)
2:1 Irradiated PBMCs (day 1, 8, 15?)
0.2:1 Irradiated Cetuximab<sup>+</sup> K562 (day 1, 8, 15?)

Assess CAR expression (GFP+) by flow cytometry

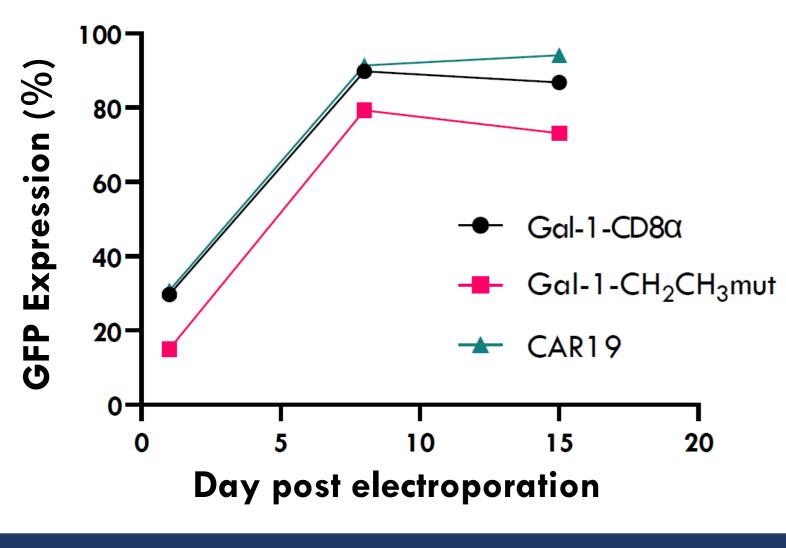
**Day 22** 



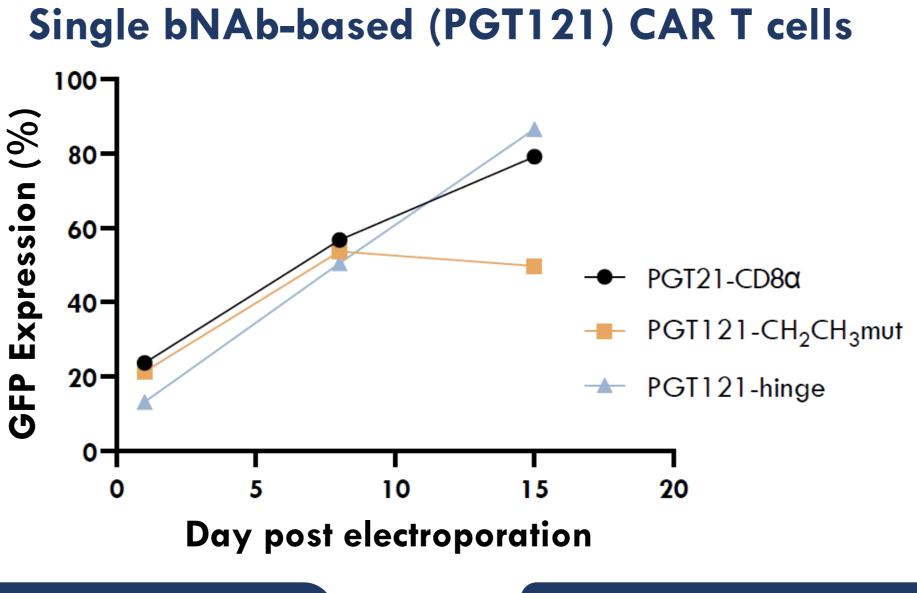


# RESULTS

## Single Gal-1 CAR T cells



# GFP expression on CD8 CAR T cells

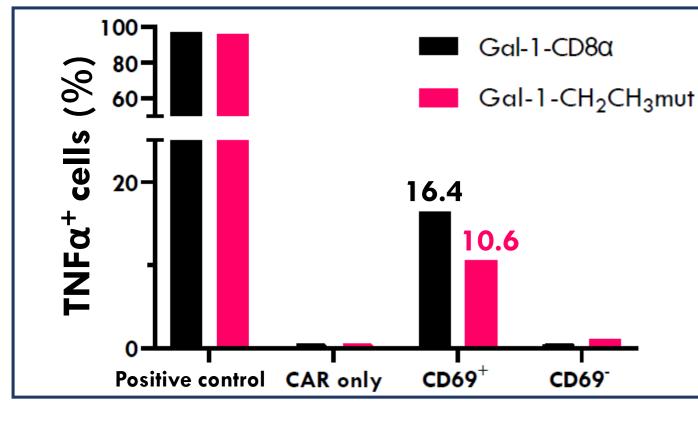


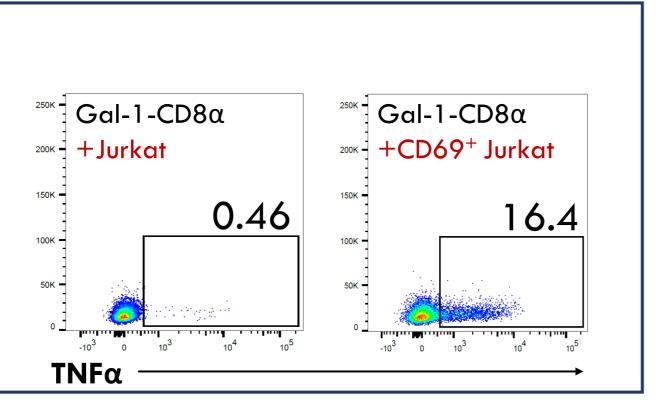
# Split TanCAR T cells Gal-1-CD8a PGT121-CD8a Split TanCAR-CD8a

Day post electroporation

# Specific target cells activate CD8 CAR T cells to express anti-viral cytokines

## Gal-1 CAR + CD69<sup>+</sup> Jurkat





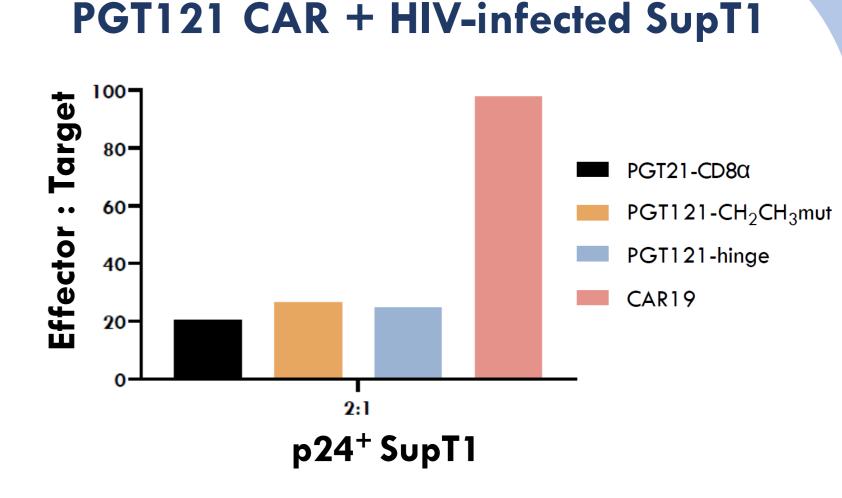
# CD8 CAR T cells exhibit cytotoxicity against targets

Gal-1 CAR + CD69<sup>+</sup> Jurkat

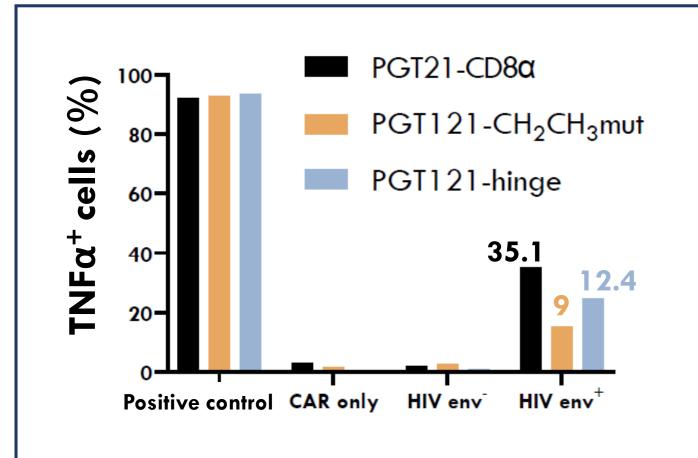
Gal-1-CD8a Gal-1-CH<sub>2</sub>CH<sub>3</sub>mut CAR19

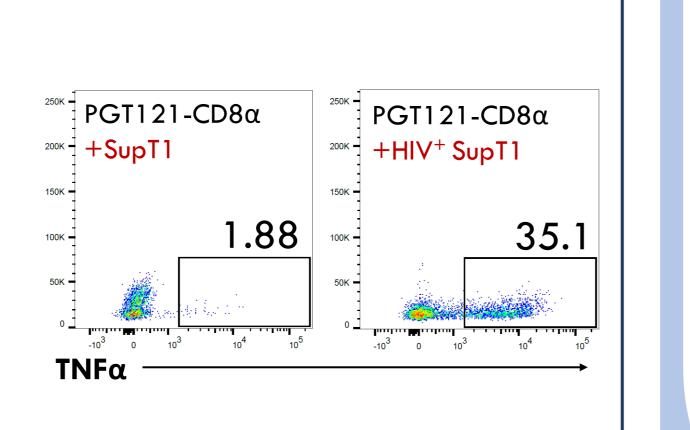
Gal-1-CD8a Gal-1-CH<sub>2</sub>CH<sub>3</sub>mut CAR19

Effector: Target

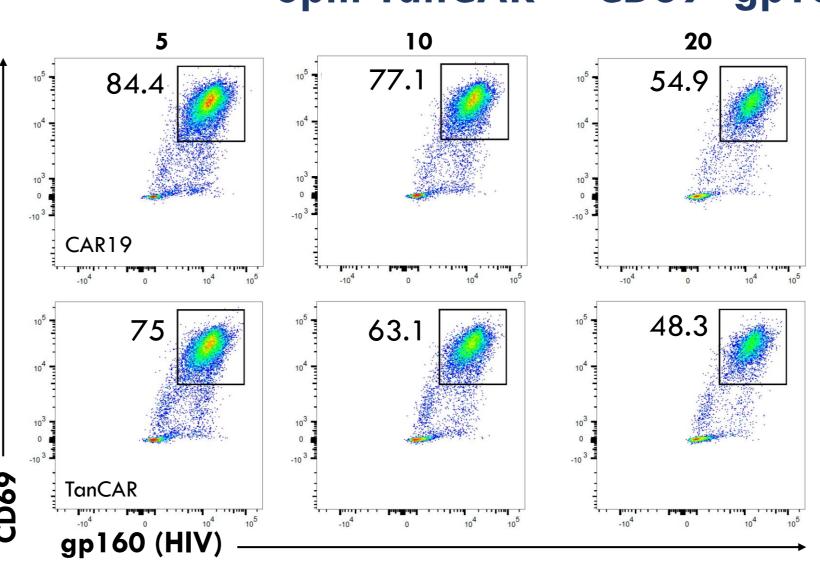


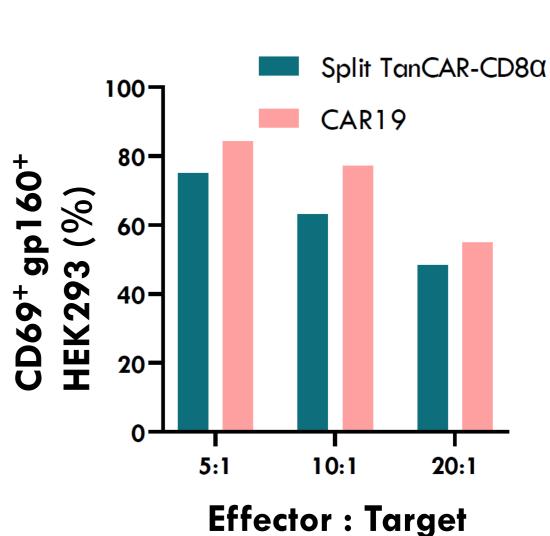
## PGT121 CAR + HIV-infected SupT1





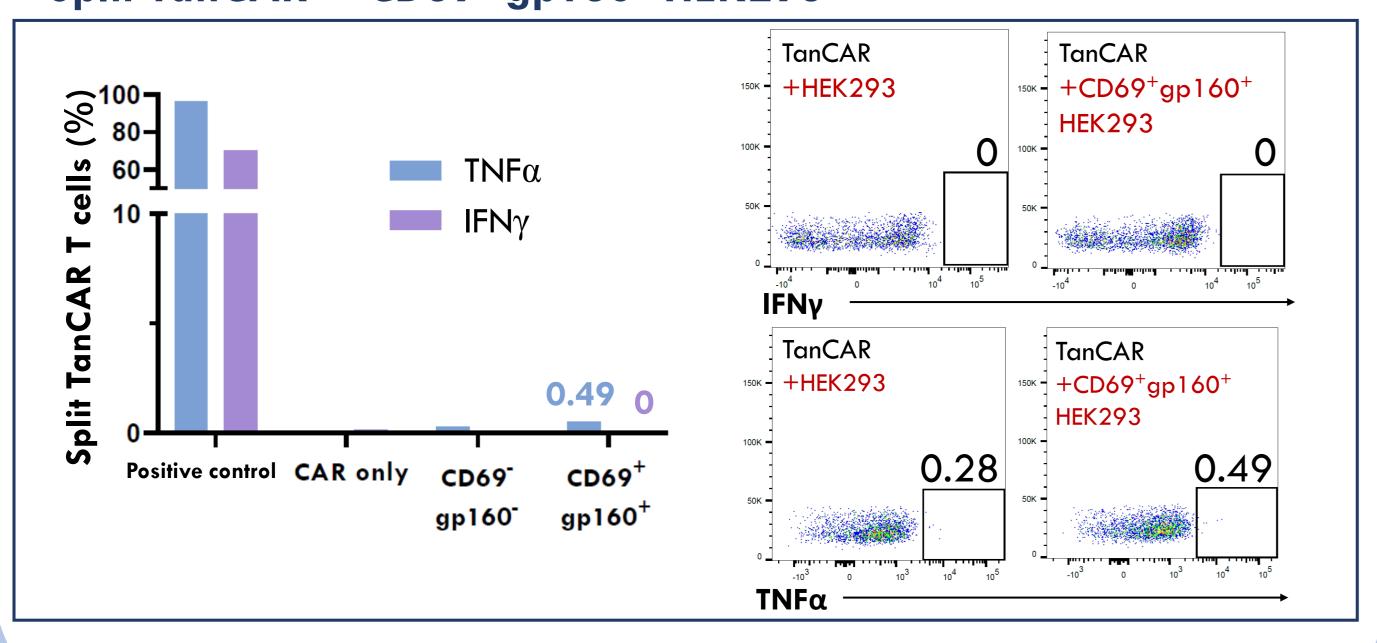
# Split TanCAR + CD69<sup>+</sup> gp160<sup>+</sup> HEK293





Flow cytometry data shows number of live target cells after co-culture with different effector to target ratio of single or split TanCAR CD8 T cells with their target cells for 6 hours compared to non-specific CAR19.

# Split TanCAR + CD69<sup>+</sup> gp160<sup>+</sup> HEK293



Flow cytometry data shows TNF $\alpha$  and IFN $\gamma$  (not shown for single CARs) expressingcells after co-culture of single or split TanCAR CD8 T cells with their target cells for 4 hours.

# CONCLUSIONS AND FUTURE DIRECTIONS

- Single CARs utilising the CD8 $\alpha$  spacer were the best in effector functions. Therefore, split TanCAR was generated using CD8 $\alpha$ .
- Split TanCAR did not exert any effector function.

**Future directions:** Investigate steric hindrance on split TanCAR T cells by generating other CAR iterations (Split or Tandem CAR)

**Significance:** Development of dual target immunotherapy may provide a novel approach in accessing HIV tissue reservoir and overcoming antibody resistance.







